

POSTER NO: DAY 1-  
PO3

## ESTIMATION OF TOTAL IRON AND TRANSFERRIN BOUND IRON (TBI) IN HUMAN SERUM USING ICP-OES AND ITS APPLICATION ON BIOEQUIVALENCE STUDIES OF IRON-SUCROSE INJECTION

Devina Bhardwaj\*, Mital Jani, Dr. ShivPrakash Rathanam

Intervein Laboratories Pvt. Ltd.,

501-503, Shapath-IV, Tower-B, S.G. Highway, Ahmedabad-380051, Gujarat, INDIA

devina@intervelabs.com



### INTRODUCTION

1.6 Billion people are affected by Iron-deficiency anemia worldwide. IV Iron therapy commonly administered to dialysis patients as an adjunct to managing anemia provides a potentially rich source for intradialytic bioactive Iron. Iron-Sucrose, Iron-dextran or ferrie glucomate complex like venofer or Iroferon is used to treat Iron deficiency primarily in patients on chronic dialysis and also in patients unable to take oral Iron supplements. After parenteral administration, the concentration of total Iron remain substantially increased. Most colorimetric analysis of Iron detect only small quantities of Iron-sucrose in serum because Iron-sucrose is stable in acidic media & Iron chelating cannot dissociate Iron from sucrose or dextran. Development of these pharmaceutical agents requires pharmacokinetic studies monitoring levels of both administered agent & transferrin bound Iron. Accurate quantitative detection of serum Iron components in the presence of Iron carbohydrates have proven problematic. Iron Sucrose present in serum tends to interfere with conventional techniques for TBI detection that utilize colorimetric or Spectrophotometry. Such techniques require reduction of ferrie ion to ferrous ion & Chelation of ferrous iron to form a colored complex. These procedures fail to accurately distinguish between Iron originating from Iron carbohydrate and Iron originating from TBI. Conventional detection techniques requiring ferrous reduction not only release Iron from TBI but also release Iron from Iron carbohydrate complex. This results in overestimation of TBI present in serum. An inaccurate measurement of TBI may cause false detection of TBI over saturation indicating existence of free Iron, a condition that may cause acute toxicity.

A need therefore exists for a method & equipment for isolation of Iron components in serum that contain Iron-Sucrose complex. Further, We also need a method for isolating and directly measuring amount of TBI and the amount of Iron carbohydrate present in the serum after administration of Iron-sucrose complex. Pharmacokinetic studies of IV Iron sucrose are complicated by background circulating Iron levels as well as need to differentiate and independently monitor Iron-sucrose and TBI. Prior to administration of product, most circulating Iron is in the form of TBI. Following IV therapy, circulating Iron is both as component of dose formulated & complexed to transferrin. The typical colorimetric assays are incapable of discriminating and accurately quantifying TBI or total Iron levels following Iron sucrose administration. The measurement of TBI requires separating & excluding free Iron and drug derived Iron from TBI.

We have developed and validated a robust and accurate method for analysis of total Iron and TBI to generate Pharmacokinetic data as component of Bioequivalence studies for generic IV. The measurement device used for total Iron & Transferrin Bound Iron was ICP-OES Iron from Iron sucrose complex. Circulating Iron-Sucrose could then be calculated as difference between total

### MATERIALS

Iron isolation, 1G/dl used as Standard was purchased from Fisher scientific while Indium Standard, 1G/dl purchased from Fluka.

Sodium Nitrate, Sodium Chloride purchased from Merck, Cysteine Hydrochloride Monohydrate purchased from Loba chemicals and Calcium Nitrate Tetrahydrate purchased from SDFCL were used for preparation of Simulated Serum.

Holo transferrin Human purchased from sigma was used for preparation of TBI QC's. Nitric acid purchased from Merck while HPLC water purchased from Rankem.

Alumina Basic Cartridges, 500mg/3 ml. Extraction cartridges purchased from Orcochem.



### EQUIPMENT

A Thermo Scientific Inductively Coupled Plasma System Equipped with Optical Emission Spectrophotometer (7000 Series) and System controlled through Qtegra Software.

#### ICP-OES Condition:

Opt. Emission	Acid
UV Exposure Time	15
UV RF Power	1150
UV Neb Gas Flow	0.5
VIS Exposure Time	5
VIS RF Power	1150
VIS Nebulizer Gas Flow	0.5
Cost Gas Flow rate	12
Auxiliary Gas Flow Rate	0.5
Iron Wavelength	259.940
Indium wavelength	325.609



### METHOD VALIDATION

Method Used for Determination of Iron and Transferrin Bound Iron in Human plasma was developed and Validated for ICP-OES.

#### TOTAL IRON

Method Validated in terms of System Suitability, Specificity, Matrix effect, Linearity, Accuracy & Precision, Extraction Efficiency, Dilution Integrity and Stability experiment.

System suitability performed by aspirating 6 replicate of higher standard.

Calibration standard used for linearity were prepared in Simulated serum.

QC's were used for Accuracy & Precision determination were prepared in Human serum and Simulated serum.

Specificity was evaluated by using 4 normal serum as well as 1 Hemolytic and Lipemic Serum.

Extraction efficiency was evaluated by comparison of Concentration of HQC, MQC and LQC level in Simulated serum as compare with concentration of HQC, MQC and LQC in Human Serum.

Dilution Integrity evaluated for dilution Factor: 2 and 5 times of High QC.

Stock Solution: Stability Performed at HQC and LQC level at -20 $\pm$ 5  $^{\circ}$ C.

Freeze thaw stability performed at HQC and LQC level in Human serum and Simulated serum at -20 $\pm$ 5  $^{\circ}$ C.

Bench Top stability performed at HQC and LQC Level in Human Serum and Simulated Serum at RT.

Long Term stability performed at HQC and LQC level in Human Serum at -20 $\pm$ 5  $^{\circ}$ C.

#### TBI

Method Validated in terms of System Suitability, Linearity, Accuracy & Precision, Extraction Efficiency, Dilution Integrity and Stability experiment.

Holo Transferrin Human was added into Human Serum.

System suitability performed by aspirating 6 replicate of higher standard.

Calibration standard used for linearity were prepared in Simulated serum.

QC's were used for Accuracy & Precision determination were prepared in Human serum.

Extraction efficiency was evaluated by comparison of Concentration of HQC, MQC and LQC level in Simulated serum as compare with concentration of HQC, MQC and LQC in Human Serum.

Dilution Integrity evaluated for dilution Factor: 2 and 5 times of High QC.

Stock Solution: Stability Performed at HQC and LQC level at -20 $\pm$ 5  $^{\circ}$ C.

Freeze thaw stability performed at HQC and LQC level in Human serum at -20 $\pm$ 5  $^{\circ}$ C.

Bench Top stability performed at HQC and LQC Level in Human Serum at RT.

### RESULTS

TOTAL IRON	
SPECIFICITY	
No significant interference is observed from concomitant analyte at the wave length of Iron.	
MATRIX EFFECT	
% CV for Mean of Matrix Factor for HQC-1.1% & LQC-0.4%	
% CV of STD Normalized Factor for HQC-1.1% & LQC-1.3%	
LINEARITY	
Method was found Linear from 25.250 $\mu$ g/dl to 200.000 $\mu$ g/dl in Simulated Serum	
Correlation coefficient > 0.98	
% Accuracy for CC Standard range: 98.2% to 105.1%	
% CV for CC Standard range: 0.6 % to 3.6 %	

HUMAN SERUM	
SIMULATED SERUM	
Intra Day Accuracy: 95.0 % to 108.1 % (H, M, L)	
Intra Day Accuracy: 102.2 % to 104.6 % (H, M, L)	
Intra Day Accuracy: 98.2 % to 116.3 % (L)	
Intra Day Accuracy: 103.2 % to 118.4 % (L)	
Intra Day Precision: 0.2 % to 0.91 % (H, M, L)	
Intra Day Precision: 0.4 % to 1.09 % (H, M, L)	
Intra Day Precision: 1.2 % to 2.90 % (L)	
Intra Day Precision: 1.21 % to 5.14 % (L)	
Inter Day Accuracy: 99.5 % to 102.6 % (H, M, L)	
Inter Day Accuracy: 99.7 % to 99.5 % (H, M, L)	
Inter Day Accuracy: 108.9 % (L)	
Inter Day Accuracy: 3.2 % to 4.5 % (H, M, L)	
Inter Day Precision: 7.0 % (L)	
Inter Day Precision: 6.9 % (L)	